Dose Dependent Morphological Effects of Alpha Lipoic Acid on Vasospastic Femoral Artery in Rats

Suçanlarda Alfa Lipoik Asidin Vazospastik Femoral Arterde Doza Bağımlı Morfolojik Etkileri

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INTRODUCTION

Cerebral vasospasm (CV) is a significant source of morbidity and mortality in patients with subarachnoid hemorrhage (SAH). Despite of many articles published each year on the topic of CV, the pathophysiology has not been clearly demonstrated and there is still no efficient treatment against this delayed effect of SAH. The exact mechanisms by which SAH induces arterial vasospasm remains to be a subject of considerable research and debate.

Alpha lipoic acid (α-LA), an organo-sulfur compound derived from octanoic acid, have the characteristics of strong natural antioxidant, penetrating blood brain barrier, no serious side effects, soluble in both fat and water, anti-inflammatory and metal chelating effects. α-LA is essential for life and aerobic metabolism. α-LA has been used nutritionally and clinically since the 1950s for a number of diseases and conditions. One of the most studied clinical uses of α-LA is the treatment of diabetes and diabetic neuropathy (15). Neuroprotective effect of α-LA has been shown in several studies on neurological diseases, such as traumatic brain injury (18), cerebral ischemia (17), seizures (6), autoimmune encephalomyelitis (10), and Alzheimer (1) and Parkinson diseases (3). The aim of the current study was to determine dose dependent effects of
α-LA on vasospastic femoral artery in a rat model by analysing the histological and morphometric changes.

**MATERIAL and METHODS**

The present experimental study was conducted under the approval of the ethics committee institute for experimental medical research, (DETAE), Istanbul Faculty of Medicine, Istanbul University. The surgical part of the study was performed in the animal laboratory of the same institute. Histopathological and morphometric analyses were carried out in the laboratory of pathology department of Sisli Etfal Education and Research Hospital. Thirty two male Sprague-Dawley rats weighing 210-260 gr, microsurgical tools and surgical microscope were used in our study. The rat femoral artery vasospasm model of Okada et al. was preferred as vasospasm model (16).

Thirty two rats were randomly separated into four groups: group I (n=8), control group; group II (n=8), vasospasm group; group III (n=8), vasospasm + intraperitoneal low dose (20 mg/kg) α-LA treatment group; and group IV (n=8), vasospasm + intraperitoneal high dose (100 mg/kg) α-LA treatment group. All rats were exposed to percutaneous intraperitoneal 60mg/kg ketamine HCl (Ketalar vial, Eczacıbaşı, Turkey) and 10 mg/kg xylazine (Rompun 2%, Bayer, Turkey) for sedation and they were located on the wood block in supine position and allowed to breathe spontaneously. While performing sterile microsurgical technique, the femoral vascular bundle was exposed in the inguinal region by a longitudinal 2 cm skin incision under surgical microscope. The femoral artery was dissected meticulously from the femoral vein and nerve without being traumatized. 1-1.5 cm of a silastic sheath was wrapped around the femoral artery and sutured. Autologous fresh tail blood of the subjects was used as whole blood. In groups II, III and IV, 0.1 ml fresh tail blood was applied directly to the femoral artery in the silastic sheath for generation of a peripheral vasospasm model. Control group I femoral arteries were covered only with a silastic sheath. As the rats in groups I and II were delivered intraperitoneal 0.3 ml/day 0.9% saline, the rats in group III were subjected to intraperitoneal 20 mg/kg/day α-LA (Thioctacid®, Gen ilac, Ankara, Turkey) and the rats in group IV were subjected to intraperitoneal 100 mg/kg/day α-LA for seven days. Rats were fed with mouse diet and kept at room temperature in separate cages for seven days. No subject required exclusion due to mortality or disease.

On day 7, rats were reanesthetized intraperitoneally with 50 mg/kg ketamine and 10 mg/kg xylazine. The silastic sheath wrapped around the femoral artery was accessed by opening the earlier surgical incisions and the femoral artery was exposed from the silastic sheath. Meanwhile, the surface of the sternum was shaved. Following sterilization with povidine iodine, the sternum was cut from the attachment points with the ribs and the thorax was explored. By opening the pericardium, puncture was applied to the left ventricle with a green-tipped injection needle. A serum set was fixated onto the end of this catheter, perfusion was performed at a standard height of 100 cm from the chest and 300 ml of 0.9% NaCl solution was delivered to the left ventricle. A mixture of 100 ml 0.03M phosphate buffer, 200 ml of 4% paraformaldehyde and 1% glutaraldehyde solution was delivered to the left ventricle. The administration of the solution was continued until spotting a clear fluid coming out of the right atrium. After perfusion fixation, right femoral artery was incised 1–1.5 cm long and removed for histopathological and morphometric analysis in all groups. After this procedure, the rats were killed by delivering a megadose of intraperitoneal ketamine. Femoral artery specimens kept in 10% formaldehyde were put into the tissue processor and treated with formalin for fixation purposes. Following dehydration with graded alcohol, it was processed in xylene stage and embedded in paraffin. Frozen paraffin sections of 5 μ thickness were obtained and deparaffinized for 1 h in the incubator. Deparaffinization was continued with xylene (three times). The sections were treated with graded alcohol, rehydrated, washed with water, and stained with hematoxylin eosin. The preparations were examined under ×100, ×200, and ×400 magnifications with an Olympus (Olympus BX7, Japan) microscope and their photographs were taken for morphometric analysis. Vascular wall thicknesses and lumen cross sectional areas were measured as unit values by the micrometer. Measurements were carried out on the photographs taken under ×40 magnification. The specimens were compared in terms of vascular lumen area and wall thickness in the morphometric analysis.

Vascular wall thicknesses and vascular lumen areas were morphometrically measured in all groups and evaluated statistically by SPSS (v. 15.0) and Kruskal-Wallis one way analysis for variance. A value of p<.05 was recognized as statistically significant.

**RESULTS**

**Histological Changes**

Femoral artery cross sections were examined under light microscope in all the four groups. Arterial wall in group I had thin and normal smooth endothelium, thin and unfolded internal elastic lamina, concentrically oriented smooth muscle cells, and the lumen was clearly observed (Figure 1). Arteries in vasospasm alone group II, under light microscopy, had significant narrowing in the lumen width, apparent thickening in the vascular wall, disruption of endothelial integrity, folding in the internal elastic lamina, and vacuolizations in the muscle layer (Figure 2).

In 20 mg/kg α-LA administered group III, light microscopy found out that femoral artery wall had a thinner and smoother endothelium than the group II, mildly folded internal elastic lamina, and concentric smooth muscle cells (Figure 3).

In 100 mg/kg α-LA administered group IV, light microscopic examination of the femoral artery showed vascular wall had a thin and smooth endothelium, very thin and mildly folded internal elastic lamina, and concentric smooth muscle cells. This appearance was similar to the control group I (Figure 4).
**Morphometric Analysis**

Comparisons across rat groups were performed after completing of measurement of the femoral artery wall thicknesses and lumen areas (Table I, II; Figure 5, 6). The lumen cross sectional area and wall thickness of all the femoral arteries were summarized as the mean and standard deviation values. Vasospasm alone group (group II) demonstrated the higher vessel wall thickness and the lower lumen area as compared with group I (p<.05; Table I, II). This was indicative of the development of a considerable vasospasm following delivery of blood to the periarterial areas compared with the control group.

Group IV showed a significant decrease in terms of mean vascular wall thickness and increase in lumen area compared with group II (p<.05; Table I, II). No significant difference was found between the mean vascular wall thickness and lumen area of groups IV and I (p>.05), while group IV showed larger lumen area than group I in measurements but it was not significant statistically. The statistical comparison of the mean vascular wall thickness between group III (low dose α-LA) and IV (high dose α-LA) displayed no significant difference (p>.05), whereas comparison of vascular lumen areas showed a significant increase in group IV (p<.05; Table III).

We also measured blood glucose levels of the subjects before starting the study and ending for histological analysis because of that α-LA has been used in the treatment of diabetes mellitus. The comparison of first and last blood glucose levels of each groups showed that the decrease of last blood glucose levels of groups 3 and 4 (α-LA administered) was statistically significant (p<.05; Table IV). The first and last blood glucose levels of groups 1 and 2 (control and vasospasm groups) was found insignificantly different (p>.05; Table IV). During the study, we were able to know that none of the subjects showed hypoglycemia.

![Figure 1: Light microscopic view of the control group (Group I; H&E, x200).](image1)

![Figure 2: Light microscopic view of the vasospasm group (Group II; H&E, x200).](image2)

![Figure 3: Light microscopic view of low dose 20 mg/kg α-LA administered group (Group III; H&E, x200).](image3)

![Figure 4: Light microscopic view of high dose 100 mg/kg α-LA administered group (Group IV; H&E, x200).](image4)
Figure 5: Comparison of mean vessel wall thickness of the four groups.

Figure 6: Comparison of vascular lumen area of the four groups.

Table I: Statistical Analysis of the Mean Vessel Wall Thickness Values of the Four Groups ± Standard Deviation (mm)

<table>
<thead>
<tr>
<th></th>
<th>Group I</th>
<th>Group II</th>
<th>Group III</th>
<th>Group IV</th>
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<tbody>
<tr>
<td>Mean</td>
<td>0.088</td>
<td>0.155</td>
<td>0.144</td>
<td>0.121</td>
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<tr>
<td>±Sd</td>
<td>0.018</td>
<td>0.044</td>
<td>0.035</td>
<td>0.021</td>
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</table>

*p<.05

Table II: Statistical Analysis of the Mean Vascular Lumen Area Values of the Four Groups ± Standard Deviation (mm²)

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<tr>
<td>Mean</td>
<td>0.369</td>
<td>0.214</td>
<td>0.224</td>
<td>0.425</td>
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<tr>
<td>±Sd</td>
<td>0.214</td>
<td>0.042</td>
<td>0.072</td>
<td>0.229</td>
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*p<.05

Table III: Statistical Analysis of the Mean Vessel Wall Thickness Values of the Groups III and IV ± Standard Deviation (mm)

<table>
<thead>
<tr>
<th></th>
<th>Group III</th>
<th>Group IV</th>
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<tbody>
<tr>
<td>Mean</td>
<td>0.144</td>
<td>0.121</td>
</tr>
<tr>
<td>±Sd</td>
<td>0.035</td>
<td>0.021</td>
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*p>.05

Table IV: Statistical Analysis of the Mean First and Last Blood Glucose Levels (BGL) of the Four Groups ± Standard Deviation (mg/dL)

<table>
<thead>
<tr>
<th></th>
<th>First BGL</th>
<th>Last BGL</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean</td>
<td>±Sd</td>
<td>Mean</td>
<td>±Sd</td>
</tr>
<tr>
<td>Group I</td>
<td>296,88</td>
<td>286,25</td>
<td>41,55</td>
</tr>
<tr>
<td>Group II</td>
<td>266,63</td>
<td>215,75</td>
<td>29,41</td>
</tr>
<tr>
<td>Group III</td>
<td>268,38</td>
<td>210,63</td>
<td>40,30</td>
</tr>
<tr>
<td>Group IV</td>
<td>297,00</td>
<td>200,88</td>
<td>26,33</td>
</tr>
</tbody>
</table>

*p<.05
DISCUSSION

Pathophysiology of cerebral vasospasm is complex and multifactorial. The oxidative stress is one of the factors which provoke vasospasm after SAH (11, 12). It is believed that oxidative stress stimulates hypertrophy and proliferation of smooth muscle cells and causes the increase of endothelial apoptosis. These changes are related to the contractile response of the vessels. There is a correlation between the development of cerebral vasospasm and increased superoxide anion levels in cerebrospinal fluid after SAH (2). Various studies of vasospasm performed on several animals e.g. rats, pigs, cats, dogs, and rabbits have been reported. It is found that ferrous chelators, eg. Ebselen, Tililazad, Nicaraven and inhibitors of the enzymes producing free radicals, reduce the vasospasm on animal SAH models (7, 8, 9, 14, 19).

We analysed the morphological evaluation of vasospasm by using femoral artery vasospasm model of Okada et al. (16). Although the response of cerebral arteries against injury is not the same as systemic arteries, clinical and experimental knowledge suggests that vasospasm is related directly to the blood volume and the time interval in which vessel wall is in contact with the whole blood (14). The silastic cuff was used to lengthen the time interval and the quantity of whole blood around the adventitia. In our study, arterial vasospasm was clearly seen in the vasospasm group comparing to the control group.

α-LA, or thiolic acid, or 1,2-dithiolane-3-pentanoic acid, is a naturally occurring dithiol compound synthesized enzymatically in the mitochondria from octanoic acid. In addition to synthesis, α-LA is also absorbed intact from dietary sources, and it transiently accumulates in many tissues. α-LA is a low molecular weighted substance that crosses the blood brain barrier and it does not have any serious side effects. Although the metabolic role of α-LA has been known for more than 50 years, its effects when given exogenously have become known recently. Several studies have provided evidence that α-LA supplementation decreases oxidative stress and restores reduced levels of other antioxidants in vivo. α-LA is an important co-factor in pyruvate dehydrogenase and α-ketoglutarate in the mitochondria. It was found that α-LA inhibits α-ketoglutarate dehydrogenase mediated generation of superoxide and hydrogen peroxide. α-LA has a variety of properties which can interfere with the pathogenesis or progression of neurodegenerative diseases (13).

α-LA is effective in treatment of diabetes, furthermore it had remarkable therapeutic potential in vascular diseases, hypertension and inflammation. Since SAH related cerebral ischemia still has an important role on mortality while prognosis of the surviving patients remains poor, extensive experimental and human studies are currently performed to build up therapeutic strategies for better protection of the cerebral tissues. Panigrafi et al. have shown that the natural thiol antioxidant α-LA is effective in improving survival and protecting the rat against reperfusion injury following cerebral ischemia (17).

The effectiveness of dihydrolipoic acid (DHLA), reduced form of α-LA, in prevention of reperfusion injury has been shown previously (4). α-LA and DHLA are capable of scavenging reactive oxygen species such as hydroxyl radicals, peroxyl radicals, superoxide, hypochlorous acid and peroxynitrite. They prevent singlet oxygen induced DNA damage, exhibit chelating activity, reduce lipid peroxidation and increase intracellular glutathione levels. Therefore α-LA and DHLA have the potency to improve SAH induced oxidative injury.

Considering a recent study of Ersahin et al. (5), α-LA has a significant neuroprotective effect mediated via enhancing the activity of endogenous antioxidant enzymes, inhibiting neutrophil accumulation and free radical generation. It was reported that α-LA treatment (100 mg/kg/day, po for 2 days) suppressed the high percentage of DNA fragmentation in the brain, while oxidative brain injury was improved. The study has determined α-LA reversed the SAH induced changes, such as increased formation of reactive oxygen radicals, increased DNA fragmentation, increased myeloperoxidase activity, depletion of brain glutathione level, increased Na⁺/ K⁺-ATPase activity, to normal significantly on experimental SAH model of rats. In the basilar arteries of the rats treated with α-LA, arterial vasospasm, similar to our histological results with α-LA (100 mg/kg/day, intraperitoneal for 7 days), was decreased providing the integrity of the endothelium.

In the current study, the dose of intraperitoneal 100 mg/kg/day α-LA decreased the vasospasm and improved the morphological changes related to damage of the vascular structures. These findings could not be seen at the dose of intraperitoneal 20 mg/kg/day α-LA. Lumen area of the vessel is especially calculated rather than lumen diameter for each groups. In the group, high dose α-LA administered after vasospasm, light microscope analysis showed that all vessels had thin and smooth endothelium, mildly folded internal elastic lamina, concentrically oriented smooth muscle cells resembling the control group. Vascular wall thickness of group IV is significantly lower than group II, and the mean lumen area of group IV is significantly higher than the groups II and III. The results obtained from the study demonstrate high dose of α-LA is efficient in a peripheral artery SAH animal model.

We measured blood glucose levels of the subjects before starting the study and ending for histological analysis because α-LA is currently being used in the treatment of diabetes mellitus. The comparison of first and last blood glucose levels of each groups showed that the amount of decrease in blood glucose levels of the groups 3 and 4 (α-LA administered) was significantly different from that of the groups 1 and 2 (control and vasospasm groups). This finding shows that the effect of the α-LA treatment on blood glucose level should not be ignored. Hypoglycemia was not seen in any group of the subjects during the study.

In addition to the literature, by the present study, the use of low and high doses of intraperitoneal α-LA has lead to support and to improve the recent knowledge on vasospasm.
CONCLUSION

α-LA was found to prevent the development of experimental peripheral vasospasm morphometrically. This is the first study to search comprehensively the dose dependent in vivo effects of α-LA on histological and morphological alterations of vasospastic arteries in the experimental rat model. As intraperitoneal administration of 20 mg/kg and 100 mg/kg α-LA was studied in vasospasm model, we have found significant difference morphologically related to the dosage. If α-LA dosage is higher, the mean vascular lumen area increases just about the value of the control group.

We believe that the knowledge obtained from our study will shed light to other experimental studies in the determination of effective doses of α-LA against vasospasm. Further studies are required for the use of α-LA in cerebral vasospasm (CV) after SAH in human before definite conclusions can be drawn.

ABBREVIATIONS


REFERENCES


