

Effect of Axonal Length on Direct Neuromuscular Neurotization: An Experimental Study

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ABSTRACT

AIM: To compare the effect of long and short axonal lengths on neuromuscular neurotization (NMN).

MATERIAL and METHODS: In this study, 18 male Wistar rats were divided into two groups. In each group, the peroneal nerve and soleus muscle were dissected. In Group1, the muscle was neurotized after the peroneal nerve was dissected until bifurcation. In Group2, the nerve was transected 1 cm proximal from the most distal site, and the muscle was neurotized with a shorter nerve than that in Group1.

RESULTS: In Group2 (0.84), the compound muscle action potential amplitude ratio was statistically higher than that in Group1 (0.42). Upon pathological examination, the cross-sectional area was statistically larger in Group2. Acetylcholinesterase enzyme levels in Groups1 and 2 were 37.73 and 45.47, respectively.

CONCLUSION: Considering that NMN with shorter axons showed better results than that with longer axons, nerve transfers using nerves as short as possible should be preferred in clinical applications.

KEYWORDS: Axon, Experimental, Neurotization, Rat

INTRODUCTION

Neuromuscular neurotization (NMN) involves direct implantation of a donor nerve into a denervated muscle and is used to treat chronic denervation and severe nerve injuries. Although many studies have been conducted on NMN, its exact mechanism remains unclear and the optimal technique has not been established. Numerous factors affect the results of a procedure. Various surgical techniques and exogenous factors have been studied to improve the outcome, but the clinical use of NMN remains unsatisfactory (2,4,5,15,18,22,23,29).

A number of changes occur in the proximal axon, nerve cell body, and myelin sheath following axonal injury (11). The proximal nerve segment can undergo apoptosis or chromatolysis. Chromatolysis, an attempt to repair the nerve,

is accompanied by an increase in the production of structural and growth cone components needed for axonal growth, i.e., tubulin, actin, and growth-associated protein 43 (12,25,28). The location of axonal injury is critical in neuronal regeneration following a proximal nerve injury, and the consequences are more severe (10).

Thus, the objective of the current study was to compare the effect of long and short axonal lengths on NMN in a rat model of muscle denervation.

MATERIAL and METHODS

Eighteen male albino Wistar rats, weighing 275–300 g, were used. All procedures were performed in accordance with the animal research committee of our medical academia (ETİK-2014/49). Intraperitoneal ketamine, 100 mg/kg, and



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intramuscular xylazine, 35 mg/kg, were used for general anesthesia in all procedures. Standard microvascular instruments and a surgical microscope were used.

Study Groups

The rats were randomly divided into two groups. In both groups, the common peroneal nerve was transected from the most distal part before branching off. In Group2, an additional 1 cm axon was excised. All rats were euthanized on the third postoperative month, and electromyographic, biochemical, and pathological analyses were performed. Contralateral legs were used as controls for electromyography (EMG).

Surgical Procedure

The posterior gluteal region was shaved, prepared, and draped under sterile conditions. Each rat was fixed on the operating tray in the prone position. The sciatic, tibial, and common peroneal nerves were explored via a posterior incision on the right side. In Group1, the peroneal nerve was transected just before bifurcating to the deep and superficial branches. In Group2, the nerve was transected from the branching site, as

in Group1. Then, a 1 cm proximal nerve segment was excised (Figure 1).

The incision was extended to the lower leg, and the soleus muscle was explored. The branches of the tibial nerve that innervate the soleus muscle were identified and confirmed with 0.5 mA nerve stimulator. The muscle was then denervated, and the common peroneal nerve was inserted into the soleus muscle. The NMN was performed in the proximal one fourth of the soleus muscle in both groups (Figure 2A, B). The skin incision was closed with interrupted 5-0 silk sutures, and the animals were returned to their cages with free access to food and water. Acetaminophen (50 mg/kg i.p.) and cefazoline (10 mg/kg i.p.) were administered for postoperative analgesia and infection prevention.

Electrophysiological Studies

Compound muscle action potentials (CMAPs) were recorded under general anesthesia for all animals at the end of the third month. After anesthesia induction, the soleus muscle and peroneal nerve were exposed, the needle was inserted into the middle portion of the muscle, and the CMAP was recorded. The same procedure was applied to the same part of the muscle and to the contralateral legs of all animals. All investigators involved in EMG procedures were blinded to the study groups. The difference in latency was calculated by subtracting the right leg latency from the left leg latency. The amplitude ratio was calculated by dividing the right leg amplitude by the left leg amplitude.

Biochemical Assays

After the EMG, the soleus muscle was excised and divided into two pieces. The proximal half of the muscle, including the NMN site, was sent to the biochemistry laboratory. Each tissue sample was homogenized with 810 μ L of 0.1 M ice-cold phosphate buffer saline (pH 7.4). The homogenates were centrifuged at 2300 g for 20 mins (4°C). Supernatants were removed and analyzed for protein and acetylcholinesterase (AChE) levels. The protein level in supernatant samples was measured by the Lowry method (17). The ELISA kit for rat AChE was purchased from YH Biosearch Laboratory (Shanghai YeHua Biological Technology Co., Ltd., Shanghai, China). ELISA assay was performed according to the manufacturer's instructions. The AChE results were divided by protein amount and given as U/g of protein.

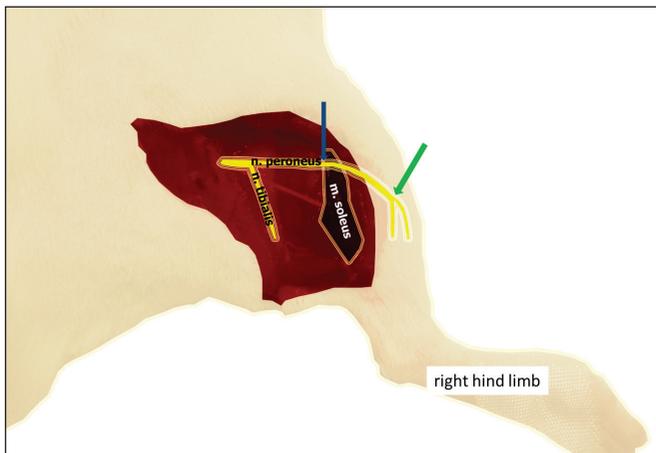


Figure 1: A schematic drawing of the experimental groups. Nerve transection was performed from the most distal part of the peroneal nerve prior to bifurcation (green arrow) in Group1. A 1 cm nerve segment was excised, and neuromuscular neurotization was performed (blue arrow) in Group2, following transection of the nerve from the most distal part.

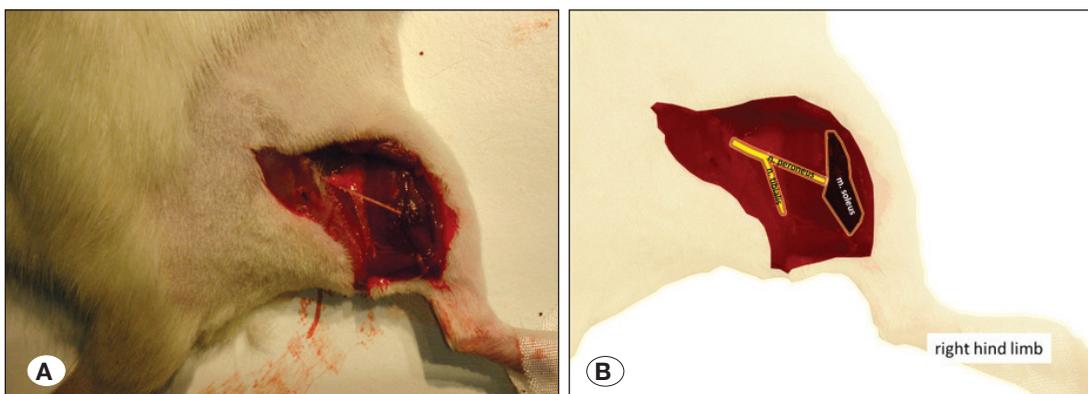


Figure 2: A depiction of the neuromuscular neurotization procedure. **A)** The distal stump was implanted in the soleus muscle following nerve transection. Implantation was carried out in the same part (1/4 proximal) of the muscle in both groups. **B)** A schematic demonstration of the surgical procedure.

Histopathological Evaluation

The distal halves of the muscle obtained for histological studies were separately fixed in a neutral formalin solution with 10% buffer for 48 hours, then dehydrated in alcohol, and embedded in paraffin blocks. Transverse sections were cut at 4 μ m thickness, deparaffinized, and then stained with Masson's trichrome and hematoxylin and eosin. The tissue samples were examined with a standard light microscope. Photomicrographs were obtained at 200 \times magnification using an Olympus BX51 light microscope and DP71 digital camera (Olympus Co., Tokyo, Japan). For each subject, 30 muscle fascicles were randomly outlined and cross-sectional areas were quantified by pixel count using ImageJ software (version 1.50i, National Institutes of Health, Bethesda, MD, USA).

Statistical Analysis

Comparisons of electrophysiological, biochemical, and pathological results among the groups were performed using t-tests for data that were normally distributed and Mann-Whitney tests for data not normally distributed. Results yielding p values less than 0.05 were considered statistically significant.

RESULTS

Two rats demised; one intraoperatively and the other due to an infection on day 5 postoperatively. They were replaced and the same surgical protocol was applied to the new animals. The weight of the animals was stable throughout the study.

Electrophysiological studies

Although there was no significant finding for the difference in the latency between both groups, a statistically significant difference was found for the amplitude ratio. At the end of the third month, for Group1, the mean difference in latency was 0.15 ± 0.06 and the mean amplitude ratio was $0.42 \pm$

0.02 . For Group2, the mean difference in latency was 0.16 ± 0.07 , and the mean amplitude ratio was 0.84 ± 0.05 . The difference in amplitude ratios between Groups1 and 2 was statistically significant ($p=0.01$). No group differences ($p>0.05$) were observed for difference in latency (Table I).

Biochemical Assays

Although AChE levels were higher in Group1, the difference between the two groups was not statistically significant ($p>0.05$). The mean AChE levels were 37.73 ± 15.43 U/g and 45.47 ± 14.19 U/g of protein for Groups1 and 2, respectively (Table II).

Histopathological evaluation

For Group1, the mean cross-sectional area contained $21,027 \pm 6,887$ pixels, whereas the mean cross-sectional area for Group2 contained $30,170 \pm 8,168$ pixels. A statistically significant group difference ($p=0.043$) was found for pixel count (Table III).

DISCUSSION

Normally innervated muscles do not permit reimplantation by another nerve. Using NMN, the original muscle nerve can be cut to change the supplying nerve so that the inserted nerve is accepted by the denervated muscle in order to generate new motor endplates (22). Although NMN is not commonly used in nerve repair, it is sometimes the only option for some patients, meaning that the need to identify ways to improve this technique is crucial.

Numerous studies have been conducted in this regard, but the exact mechanism underlying NMN is not fully understood (2,4,5,15,18,22,23,29). Several aspects of the procedure, including the source of the axon, neurotization site, depth of the neurotized axons, number of neurotized fascicles, denervation duration prior to neurotization, and

Table I: Mean Difference in Latency and Mean Amplitude Ratios of Compound Motor Action Potential in Experimental Groups at the end of the Third Month

	Group1	Group2	p
Mean difference of latencies (msec)	0.15 ± 0.05	0.16 ± 0.08	0.98
Mean amplitude ratios (mV)	0.42 ± 0.10	0.84 ± 0.12	0.01

Table II: Mean Acetylcholinesterase Levels in Neuromuscular Junctions in Experimental Groups at the end of the Third Month

	Group1	Group2	p
Mean AchE levels (U/g protein)	37.73 ± 15.43	45.47 ± 14.19	0.348

Table III: The Mean Cross-Sectional Area was Calculated (Pixel Count) by imageJ Software. After Standard Photomicrographs were Taken, 30 Fascicles were Outlined Using Software and the Mean Area of the Fascicules was Calculated

	Group1 (pixels)	Group2 (pixels)	p
Mean cross-sectional area in pixels	21027.06 ± 6887	30170.00 ± 8168	0.043

neurotransmitter effects have been examined in previous studies (1,3,6,8,13–16,19,21,27,30,31). However, the effect of axonal length on NMN outcomes has not yet been studied, and a guidance model has not been created nor established for this purpose. To address the lack of data, an animal model was designed in which the axonal length could be changed by trimming the distal stump of the peroneal nerve. Rats of similar weight were chosen to standardize the nerve length. The nerve was implanted in the soleus muscle, which was previously described and used in NMN, owing to its ideal morphological properties (7,23).

The lesion level is the primary factor affecting the outcome of end-to-end nerve repair. Poor results have been attributed to proximal lesions, with evidence to support this assumption (9,20,24,26). Various factors explain the difference in outcomes between proximal and distal lesions. For example, the nerve cell faces severe consequences after losing a large percentage of its volume following proximal injury. A long period is needed for the nerve to regenerate and reach its end organ with this type of injury. Another possibility is that proximal tributaries siphon off the regenerating axons, thereby decreasing the number of axons growing distally. A longer duration between injury and repair may reduce the functionality of the distal pathway and that of the end organs through scarring and atrophy. However, these mechanisms do not relate to the regeneration capacity of the peripheral nerve cell.

Metabolic activity increases in the neuron cell body after axotomy, and protein production begins to regenerate the distal stump of the axon (11). Thus, the current study hypothesized that the distance between the axon cell body and the distal axonal stump would affect regeneration and neurotization capacity via metabolic activity.

Electrophysiological, biochemical, and histological tests were performed to evaluate muscle reinnervation. EMG was carried out to facilitate a functional evaluation. An increased amplitude was observed in Group2, indicating enhanced axon sprouting from the distal stump. Other than the amplitude, a statistically significant difference was not found between the two groups. Latency is reflective of injury to the myelin sheath. A difference was not observed between the groups in terms of the myelin sheath status following the intervention in the current study.

AChE levels, a catalytic enzyme that is secreted from the postsynaptic membrane into the synaptic space, were determined to evaluate the new neuromuscular plates. AChE is mainly located at the neuromuscular junction, and its level correlates with the number of the neuromuscular junctions. In the current study, although a statistically significant difference between the groups was not found in AChE levels, they were found higher in Group2, potentially indicative of enhanced healing at the neuromuscular junction and of the presence of shorter axons. In line with this finding, the biochemical, pathological, and EMG results were more severe in Group1, while less muscular atrophy was observed in Group2.

The current study has some limitations. The number of animals included in the experiment and the study duration were limited

by the ethical committee. Accordingly, only a few rodents were included, and the time available for regeneration was checked. Besides, the use of a larger animal (i.e., a monkey) is warranted as a longer nerve length would better mimic a clinical situation than a rat nerve, but this was not possible in the current study.

■ CONCLUSION

Shorter axonal length was demonstrated to improve NMN healing in this rat model. To the best of our knowledge, this has not previously been achieved. When surgeons need to perform NMN (nerve transfer in congenital disorders or as a result of trauma), the use of shorter axons could yield results better than those of longer axons. Moreover, with this animal model, further research into NMN pathophysiology using different variables is warranted.

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